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# Allele-specific demethylation at an imprinted mammalian promoter

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## ABSTRACT

A screen for imprinted genes on mouse Chromosome 7 recently identified *Inpp5f\_v2*, a paternally expressed retrogene lying within an intron of *Inpp5f*. Here, we identify a novel paternally expressed variant of the *Inpp5f* gene (*Inpp5f\_v3*) that shows a number of unusual features. *Inpp5f\_v3* initiates from a CpG-rich repeat region adjoining two B1 elements, despite previous reports that SINEs are generally excluded from imprinted promoters. Accordingly, we find that the *Inpp5f\_v3* promoter acquires methylation around the time of implantation, when many repeat families undergo *de novo* epigenetic silencing. Methylation is then lost specifically on the paternally derived allele during the latter stages of embryonic development, resulting in imprinted transcriptional activation on the demethylated allele. Methylation analyses in embryos lacking maternal methylation imprints suggest that the primary imprinting mark resides within an intronic CpG island ~1 kb downstream of the *Inpp5f\_v3* transcriptional start site. These data support the hypothesis that SINEs can influence gene expression by attracting *de novo* methylation during development, a property likely to explain their exclusion from other imprinted promoters.

## INTRODUCTION

Genomic imprinting in mammals results in a small number of genes being silenced on one allele, depending on the gender of the parent from which the allele is inherited. Imprinted genes typically occur in clusters, in regions of the genome containing allele-specific DNA methylation marks which are established in the germ line. These germ-line methylation imprints can subsequently act after fertilization to establish allele-specific epigenetic asymmetry at the promoters of adjacent genes (1,2).

Mutations in members of the *de novo* methyltransferase gene family lead to disruptions in imprinted gene expression and to retrotransposon activation (3,4), suggesting that the two processes are controlled by a common mechanism (5). *Dnmt3l* encodes a regulatory protein that stimulates *de novo* methylation by *Dnmt3a* and *Dnmt3b*, but lacks the catalytic motifs necessary for methyltransferase activity. Male mice lacking functional copies of the *Dnmt3l* gene are sterile due to meiotic arrest, which is associated with the upregulation of endogenous retrotransposons (3). Females carrying null mutations in the *Dnmt3l* gene fail to establish imprinted methylation marks during oogenesis, but show no obvious effects on retrotransposon activity (6).

There is evidence to suggest that at least five known imprinted domains originated from retrotransposition events that generated novel genes during mammalian evolution. The imprinted *Peg10* gene is essential for placental development (7), and is derived from a sushi-type LTR retrotransposon that originated in a common ancestor of placental mammals (8,9). We recently reported the identification of a group of four retrotransposed gene copies (retrogenes), each of which originated from X-linked source genes and generated elements that form germ-line differentially methylated regions (gDMRs) in the mouse genome (10). Parent-of-origin effects on DNA methylation have also been extensively studied at the *agouti* locus, where stochastic methylation at an IAP insertion generates coat colour variegation (11).

Two independent studies have reported that short interspersed nuclear element (SINE) repeats are significantly underrepresented at imprinted promoters (12,13). It has been suggested that this may reflect selection against insertions within imprinted regions due to the ability of SINEs to disrupt local methylation patterns. Imprinted genes would be particularly susceptible to this epigenetic disruption due to their dependence on allele-specific CpG methylation for normal expression levels (12). However, detailed developmental methylation analyses in the region of individual SINE repeats have not been reported.

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Here, we identify a novel imprinted transcript variant of the murine *Inpp5f* gene (*Inpp5f\_v3*) which initiates from a CpG-rich region adjacent to inverted B1 SINE repeats. The promoter is situated 1 kb upstream from an imprinted and retrotransposed copy of the X-linked gene, *Tmem114a*. Both imprinted transcripts are expressed only from paternally derived alleles. A developmental analysis of CpG methylation at the two imprinted promoters suggests that the CpG island associated with the X-derived retrogene carries the primary imprint mark at the locus. The upstream repeat region undergoes *de novo* methylation at around the time of implantation, before undergoing demethylation specifically on the paternal allele during the latter stages of embryonic development. Allele-specific demethylation is a common feature of genomic imprinting in plants (14), but has not, to our knowledge, previously been reported in mammals.

## MATERIALS AND METHODS

### Tissue sources

To obtain oocytes and morulae, 4-week-old C57BL/6J (B6) females were superovulated using procedures described in (15). For oocyte collection, ovaries were dissected and cumulus clumps were flushed into hyaluronidase solution (300 µg/ml; Sigma Cat No. H4272) and left for 10 min at room temperature. To remove cumulus cells, oocytes were passed through a series of 100 µl drops of PBS in a glass Petri dish using a fine bore Pasteur pipette, and then collected in aliquots of 50 in 10 µl TE buffer. Morulae were obtained from females at embryonic day (E)3.5, and collected in aliquots of eight. Over 90% were at the 16 cell stage. E8.5 embryos were collected in pools of five for RNA purification, and individually for the isolation of genomic DNA. Embryos and tissues collected at later stages were not pooled.

The *Dnmt3l* mutant allele was generated on a 129/SvJ background. E8.5 embryos were obtained from crosses of *Dnmt3l* homozygous null mothers and wild-type (wt) cast fathers as previously described (6). The *Dnmt1* mutant allele was generated on a 129/SvJ background, and homozygous mutant, heterozygous mutant and wt E8.5 embryos were generated as described previously (16). Neonatal brains from mice carrying uniparental duplications for Chromosome 7 [proximal to the T65H breakpoint; (17)] were generated as described (Beechey *et al.*, 2004). All tissues were flash-frozen in liquid nitrogen and stored at -80°C.

### Expression studies

RNA was prepared using the RNeasy kit (Qiagen) according to the manufacturer's standard protocol, and then quantified using an Agilent 2100 Bioanalyser. cDNA was prepared from 1 µg of total RNA using the superscript II cDNA synthesis kit (Invitrogen). RT-PCR was conducted using Reddymix PCR mix (Abgene), under the following cycling conditions: 95°C for 5 min, followed by *n* cycles of 94°C for 30 s, 55°C for 30 s and 72°C for 1 min. For the amplification of *Inpp5f*, *Inpp5f\_v2*, *Bag3* and *111007A13Rik*, *n* = 32 cycles. For *Inpp5f\_v3*, *n* = 33 cycles.

In every case, control reactions were performed using cDNA prepared in the absence of reverse transcriptase, to eliminate the possibility of genomic DNA contamination. RT-PCR primer sequences are listed in Table S1. For the allele-specific RT-PCR sequencing assays, PCR products were purified using the ExoSapIT reagent (USB) and sequenced using standard ABI sequencing technology (BigDye v3.1, Applied Biosystems). RACE (rapid amplification of cDNA ends)-ready cDNA was generated using the SMART RACE cDNA kit (Clontech Cat. No. 634914) according to the manufacturer's standard protocol, using 1 µg total RNA per reaction. Touchdown PCR was performed using reagents supplied with the Advantage 2 PCR kit (Clontech Cat. No. 639207) according to the manufacturer's instructions.

### Bisulphite mutagenesis

One microgram of genomic DNA in 30 µl H<sub>2</sub>O was denatured by adding 5.3 µl 2 M NaOH (0.3 M final concentration) followed by incubation at 37°C for 15 min. Sodium metabisulphite measuring 2.1 g was dissolved in 2.5 ml of H<sub>2</sub>O, 720 µl of 2 M NaOH and 350 µl of 0.1 M hydroquinone prior to heating at 37°C for 20 min. Four hundred microlitres of this solution was added to the denatured DNA and the mixture was overlaid with 100 µl light mineral oil. Tubes were incubated in the dark for 5 h at 55°C and mineral oil was removed by serial transfer through eppendorf tubes. DNA was purified using the QiaEX II kit (Qiagen Cat No. 20021) and care was taken to shield the bisulphite-treated DNA from the light. DNA was recovered in 20 µl H<sub>2</sub>O, and desulphonated by the addition of 3.5 µl NaOH (0.3 M final concentration) followed by incubation at 37°C for 15 min. DNA was recovered by ethanol precipitation and re-suspended in 50 µl H<sub>2</sub>O.

Due to the low quantity of material available, aliquots of oocytes (50) and morulae (8) were converted using the agarose bead technique (18,19). Two percent LMP agarose was melted in a 65°C water bath, and added to the oocytes/embryos at a final concentration of 1.6%. The tubes were flicked to mix the contents and spun briefly in a benchtop centrifuge. Tubes were placed on ice for 5 min to solidify the agarose, which was overlaid with 100 µl of cold mineral oil. Four hundred microlitres of this solution (10 mM Tris-HCl pH 7.6, 10 mM EDTA, 1% SDS) was added, and 4 µl proteinase K (100 mg/ml, Sigma), prior to incubation for 15 h at 50°C. Tubes were placed on ice to resolidify the agarose and then the lysis buffer and mineral oil were removed. Agarose beads were washed for 3 × 15 min in TE buffer, 2 × 15 min in 0.3 M NaOH then finally 1 × 15 min in 0.1 M NaOH. Sodium metabisulphite/hydroquinone solution was prepared as described above and added to the agarose beads, and the mixture was overlaid with mineral oil. After incubation in the dark at 55°C for 5 h, solutions were removed and the beads were washed for 5 × 15 min in TE buffer. After the last wash, DNA was desulphonated by adding 500 µl 0.2 M NaOH and incubating at 37°C for 15 min. NaOH was then neutralized by adding 100 µl 1 M HCl and leaving to stand for 5 min. Beads were finally washed for 2 × 10 min

in H<sub>2</sub>O and all liquid was removed. The dry beads were heated at 70°C for 5 min and flicked gently to mix, then at 80°C for a further 5 min or until the agarose had melted completely. Agarose from each bead was split into two separate tubes, and used directly as a template for the subsequent PCR step. Regions 1 and 2 were amplified in nested PCR reactions, in which 2 µl of the first round reaction were carried forward to the second round. Cycling conditions were identical for each round of amplification: 95°C for 5 min, followed by 35 cycles of 94°C for 30 s, 55°C for 30 s and 72°C for 1 min. Bisulphite PCR primers were designed to be specific for the same strand from which transcription occurs, and are listed in Table S2.

### DNA sequence analysis

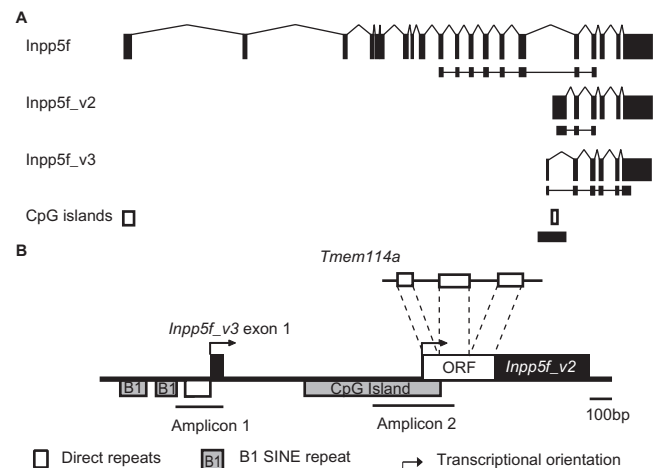
CpG islands were defined according to the criteria proposed by Gardiner-Garden and Frommer (20): >200bp, >50% C + G and an observed/expected ratio of CpG dinucleotides of >0.6 when compared to the total proportion of G + C in the same DNA segment. All repeat elements were identified using the repeatmasker tool (<http://www.repeatmasker.org/>), which is integrated into the UCSC genome browser (<http://genome.ucsc.edu/>). The region of X/autosome homology was determined using the self-chain track (Kent *et al.*, 2003) on the UCSC genome browser (<http://genome.ucsc.edu/>), which is currently available only for mouse genome build v34.

All sequence files generated by the ABI 3730xl machine were analysed using the Sequencer software package (GeneCodes). For allele-specific RT-PCR sequencing assays, the relative height of the sequence trace corresponding to each parental allele of a SNP gives an indication of the relative contribution of maternally and paternally derived alleles to the total transcript pool. For the analysis of bisulphite PCR data, strands with thymine nucleotides at positions occupied by cytosine in unconverted DNA were presumed to have been unmethylated prior to bisulphite conversion. Cytosines in CpG dinucleotides that were unconverted following treatment were presumed to have originally been methylated. Where F1 hybrid DNA was used, the parental origin of each strand was determined from a SNP within the PCR product.

## RESULTS

### The *Inpp5f* transcription unit

Recent evidence suggests that a number of mammalian imprinted domains were formed by the retrotransposition of X-linked genes into autosomal regions, generating sequences that undergo methylation in the maternal germ line (9,10). In order to gain insights into the regulation of imprinting at this class of imprinted domain, we chose to study the murine *Inpp5f\_v2* locus, searching for additional imprinted transcripts and putative regulatory elements. *Inpp5f\_v2* is a paternally expressed transcript variant of the murine *Inpp5f* gene and is transcribed at high levels in the developing nervous system (21). The unique first exon of the imprinted isoform contains a retrotransposed ORF



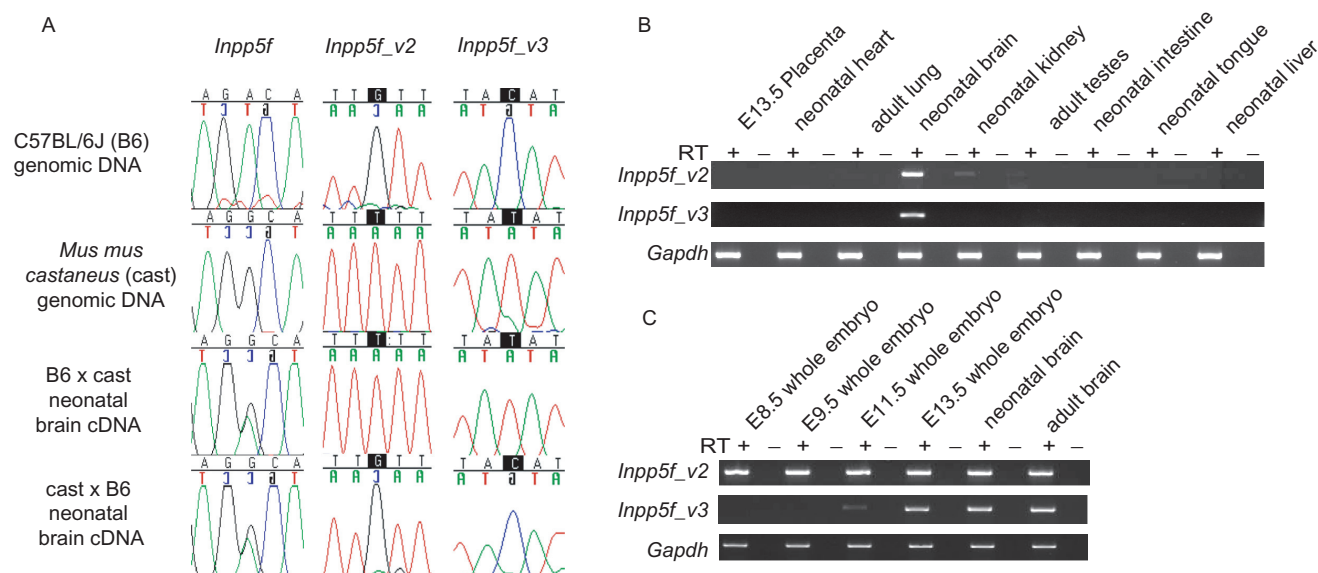
**Figure 1.** Genomic and transcriptional organization at the *Inpp5f* locus. (A) Exonic structure of *Inpp5f*, *Inpp5f\_v2* and *Inpp5f\_v3*. Exons are black rectangles and splice patterns are indicated. The RT-PCR products for the allele-specific assays in Figure 2 are indicated below each transcript. The positions of two CpG islands are shown at the bottom, and the region enlarged in panel B is indicated by a thick horizontal line. (B) Genomic features within *Inpp5f* intron fifteen. The ORF within the first exon of *Inpp5f\_v2* is a retrotransposed duplicate of the X-linked *Tmem114a* gene. The two internal promoters are represented by arrows. The region upstream of *Inpp5f\_v3* is rich in repeats, which are examined further in Figure 3. Amplicon 1 and Amplicon 2 denote the regions within which DNA methylation is examined in Figure 4.

originating from the X-linked *Tmem114a* gene, which is spliced onto five downstream exons of the non-imprinted *Inpp5f* gene (Figure 1A and B). A CpG island overlapping the retrotransposed sequence undergoes sexually dimorphic patterns of DNA methylation during gametogenesis, with maternal allele-specific methylation established during oogenesis (10). EST evidence suggests the presence of an additional promoter ~1 kb upstream of *Inpp5f\_v2*, generating a transcript variant of *Inpp5f* that has not previously been described (*Inpp5f\_v3*). RT-PCR assays demonstrate that *Inpp5f\_v3* contains a unique first exon which, like the first exon of *Inpp5f\_v2*, is spliced onto the five downstream exons of *Inpp5f* (Figure 1A). The unique first exon of *Inpp5f\_v3* lacks putative translational start codons. However, a number of in-frame start codons occur within the five exons that are shared with *Inpp5f*, potentially giving rise to C-terminal *Inpp5f* polypeptides.

### Allele-specific expression of *Inpp5f* isoforms

To assess the allele-specific expression of *Inpp5f\_v3* transcripts, RT-PCR sequencing assays were performed in neonatal brain cDNA from F1 hybrids of the C57BL/6J and *Mus musculus castaneus* sub-species. Like *Inpp5f\_v2* (21), *Inpp5f\_v3* is expressed exclusively from the paternally derived allele (Figure 2A). These findings were confirmed by the lack of *Inpp5f\_v3* expression in neonatal brains from mice carrying maternally inherited uniparental duplications for the proximal portion of Chromosome 7 (Figure S1). The 20 exon *Inpp5f* transcript is expressed from both parental alleles, as previously reported (21).





**Figure 2.** (A) Allele-specific expression of *Inpp5f* transcript variants, assessed by RT-PCR sequencing assays. PCR primers were designed to specifically amplify each transcript (Figure 1A) over regions containing SNPs between two inbred strains of mice: C57BL/6J (B6) and *Mus mus castaneus* (cast). Allele-specific expression can be determined in F1 hybrids by the relative height of the sequence trace corresponding to each parental allele at the polymorphic site. The strain of the mother is given first in each cross. (B) Tissue-specific expression of *Inpp5f\_v2* transcript variants, assessed by qualitative RT-PCR. All primers were designed to span introns (Figure 1A), and each reaction was conducted using cDNA prepared in the presence (RT+) or absence (RT-) of reverse transcriptase, to eliminate the possibility of template contamination. cDNA was prepared from 1 µg of RNA extracted from B6 × cast F1 hybrid tissues. Adult material was prepared from animals sacrificed at 6 weeks. In every case, *Inpp5f\_v2* and *Inpp5f\_v3* underwent 33 cycles of amplification, whereas *Gapdh* was amplified for 30 cycles. (C) Developmental expression screen for *Inpp5f\_v2* and *Inpp5f\_v3* transcript variants under the same experimental conditions as panel B.

The two genes flanking *Inpp5f* in the 5' (*Bag3*) and 3' (*1110007A13Rik*) direction were examined for evidence of imprinting. In each case, both parental alleles were expressed at approximately equal levels in cDNA from neonatal brain, E13.5 whole embryo and E13.5 placenta (Figure S2). These data suggest that imprinting at this locus is limited to within the *Inpp5f* transcriptional unit, but cannot exclude the imprinting of adjacent genes in a highly tissue-specific manner.

RT-PCR expression screens were conducted to determine the tissue-specific expression profile of *Inpp5f\_v3*. It has previously been shown that *Inpp5f\_v2* is expressed primarily in neural tissues (21). RT-PCR assays confirmed this finding, with expression also detected in adult testes and kidney (Figure 2B). *Inpp5f\_v3* transcripts were detected only in brain (Figure 2B). Further RT-PCR analyses were conducted in cDNA from whole embryo and brain tissues at various developmental stages. *Inpp5f\_v2* transcripts were detected in every cDNA sample assessed, indicating that expression initiates prior to E8.5 (Figure 2C). No *Inpp5f\_v3* transcripts were detected at E8.5, but expression was first detected at E11.5 and continued into adulthood (Figure 2C).

### The *Inpp5f\_v3* promoter

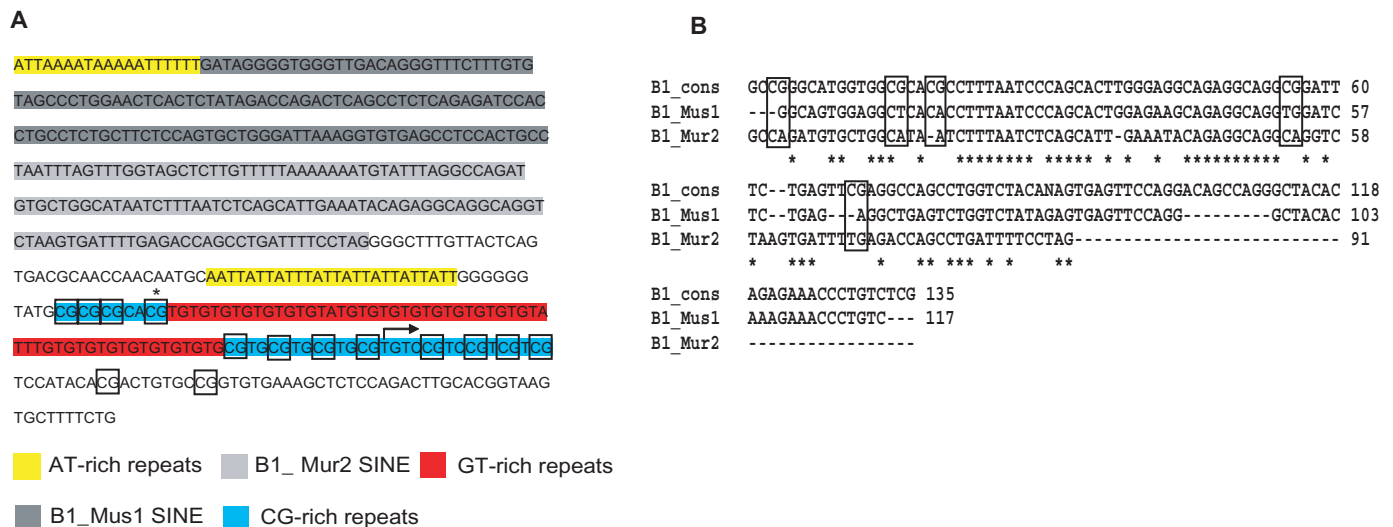
EST evidence suggests that *Inpp5f\_v3* initiates from within a CpG-rich simple repeat ~150 bp downstream of two B1 SINEs (Figure 3A). Attempts to identify the transcriptional start site by 5' RACE were unsuccessful, likely due to the CpG- and repeat-rich nature of the

5' region (Figure 3A). RT-PCR using a forward primer upstream of the GT simple repeats but downstream of the B1 SINEs fails to generate a product, whereas products are readily amplified when the forward primer overlaps the 3' terminus of the CG repeats (data not shown). This suggests that *Inpp5f\_v3* initiates from within the simple repeat region.

The SINEs belong to the Mus1 and Mur2 clades of the B1 family, and consist of 117 and 91 bp, respectively. The full-length B1 consensus sequence defined in (22) consists of 135 bp (Figure 3B). The Mus1 repeat shows 10.3% divergence from the consensus sequence annotated by repeatmasker ([www.repeatmasker.org](http://www.repeatmasker.org)), whereas the Mur2 element shows 27.5% divergence. Full-length B1 elements consist of ~135 bp, making it unlikely that either of the repeats upstream of *Inpp5f\_v3* are replication competent. Within the aligning region, the Mus1 and Mur2 elements exhibit 68% nucleotide homology with one another. The elements are arranged in an inverted 3'–5':5'–3' orientation with respect to *Inpp5f\_v3*. Although several CpG dinucleotides are present in the B1 consensus sequence (Figure 3B), all have been lost in the Mus1 and Mur2 sequences under discussion, presumably due to the spontaneous deamination of methylated cytosines to thymine nucleotides.

### DNA methylation at the *Inpp5f\_v2* and *Inpp5f\_v3* promoters

Both imprinted promoters are associated with clusters of CpG dinucleotides, for which the methylation profile was



**Figure 3.** (A) Sequence of the region upstream of and including *Inpp5f\_v3* exon one in the B6 genome. Five distinct classes of repeat, as defined by RepeatMasker ([www.repeatmasker.org](http://www.repeatmasker.org)), are shown. DNA methylation at individual CpG dinucleotides (boxed) is assessed in Figure 4. The asterisk denotes a CpG that is polymorphic between B6 and cast, and was thus excluded from methylation analyses. The furthest 5' point to which ESTs corresponding to *Inpp5f\_v3* extend is indicated by an arrow. (B) Clustalw alignment showing sequence homology between the inverted B1 SINE repeats upstream of *Inpp5f\_v3* exon 1 (B1\_Mus1 and B1\_Mur2) and the consensus sequence for the B1 family [B1\_cons; (22)]. CpG dinucleotides in the B1 consensus sequence are boxed. Note that all five consensus CpGs have been lost in the Mus1 and Mur2 sequences shown.

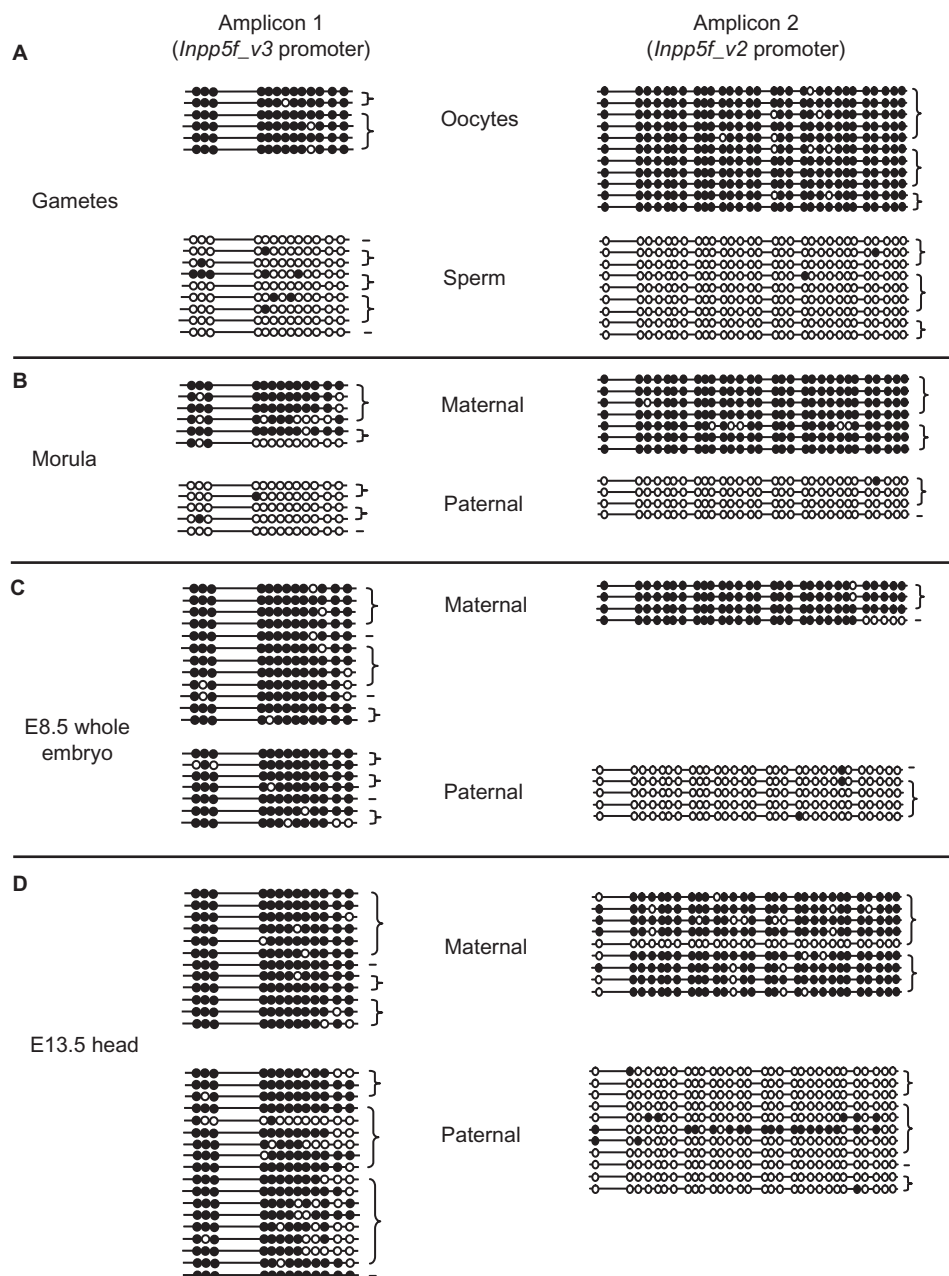
### Tissue-specific methylation differences at the *Inpp5f* v3 promoter

*Inpp5f\_v3* is expressed in the brain, but silent in all other tissues tested (Figure 2B). To determine whether tissue-specific differences in DNA methylation could contribute to the tissue-specific expression profile of this transcript, bisulphite-modified DNA from neonatal brain and heart

was sequenced over the *Inpp5f\_v3* promoter region (Figure 5). Allele-specific differences were observed in both the expressing and non-expressing tissue, but the difference was significantly less pronounced in heart than in brain (77% versus 51%; Fisher's Exact  $P < 10^{-5}$ ). Thus, the *Inpp5f\_v3* promoter shows differences in methylation both between alleles and between tissues. Tissue-specific variation in CpG methylation levels has been reported previously at the imprinted *Igf2* locus (23).

### Activation of *Inpp5f\_v3* in *Dnmt1* mutants

*Dnmt1* is responsible for the faithful propagation (or ‘maintenance’) of CpG methylation patterns during DNA replication (24). To establish whether DNA methylation is required for the silencing of *Inpp5f\_v3* at E8.5, expression was assessed in embryos that were homozygous or heterozygous for null mutations in *Dnmt1* (16), and stage-matched wt controls. Qualitative RT-PCR showed expression in *Dnmt1*<sup>-/-</sup> embryos, with a weak product also generated from *Dnmt1*<sup>+/-</sup> cDNA (Figure 6A). No expression was detected in wt E8.5 embryos. These data indicate that all of the *trans*-acting regulatory factors required for the expression of *Inpp5f\_v3* are present at E8.5, and that maintenance methylation is sufficient to silence the repeat-rich promoter. Demethylation of the paternal allele later in development (Figures 4D and 5) may therefore be sufficient for imprinted transcriptional activation in neural tissues. However, the lack of *Inpp5f\_v3* expression in heart tissue (Figure 2B), which also undergoes some degree of paternal allele demethylation (Figure 5B), indicates that neural-specific transcription factors also play a role.

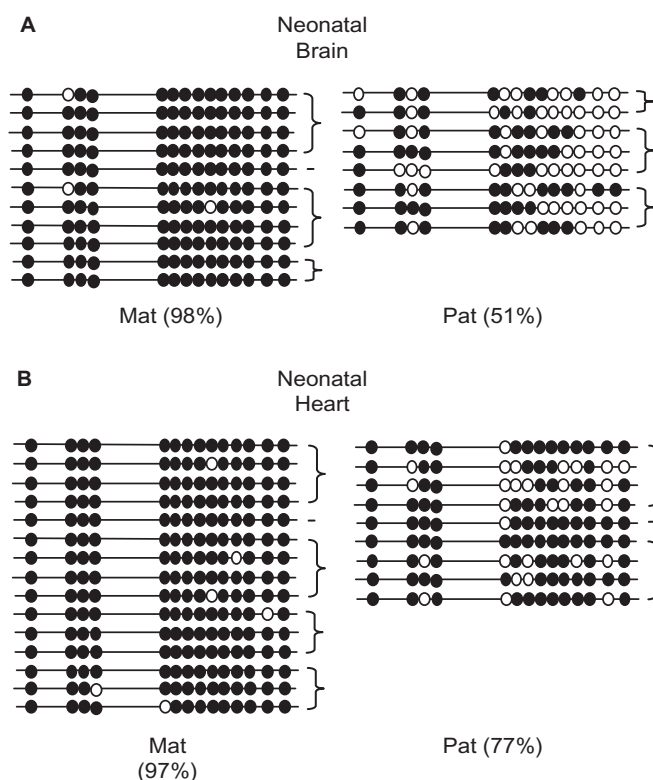


**Figure 4.** Allele-specific CpG methylation at the two imprinted promoter regions shown in Figure 1B. At embryonic stages, the parental origin of each strand was determined on the basis of SNPs between the B6 and cast alleles. Germ cell material was from purebred B6 adults. Horizontal lines represent individual strands of DNA and circles depict CpG dinucleotides. Closed circles are methylated CpGs, open circles are unmethylated. Strands derived from the same PCR amplification are connected by brace symbols to the right of the figure. The germ cell data for *Inpp5f\_v2* have been published previously (10). (A) Gametes, (B) 16 cell morulae, (C) E8.5 whole embryo and (D) E13.5 head.

#### Loss of methylation at *Inpp5f\_v2*, but not *Inpp5f\_v3*, in maternal imprint-free embryos

*Dnmt3l*<sup>-/-</sup> mothers fail to establish germ-line methylation imprints, and their heterozygous offspring die before midgestation showing disrupted expression of maternally imprinted genes (6). Maternal imprint-free embryos were generated by crossing *Dnmt3l*<sup>-/-</sup> females with *Mus mus castaneus* males, and were collected at E8.5. Bisulphite-modified genomic DNA was sequenced over the CpG-rich promoter regions of *Inpp5f\_v2* and *Inpp5f\_v3*.

The *Inpp5f\_v2* promoter region shows a lack of maternal allele-specific methylation in the maternal imprint-free embryos (Figure 6B), which is associated with reactivation of the normally silent maternal allele (Figure 6C). Surprisingly, the repeat-rich *Inpp5f\_v3* promoter is methylated on both alleles in maternal imprint-free embryos (Figure 6B). This region is methylated on both alleles in wt embryos at E8.5 (Figure 4C) and is hence unaffected by maternal *Dnmt3l* deficiency. Taken together, these data suggest that the epigenetic mark conferring memory of



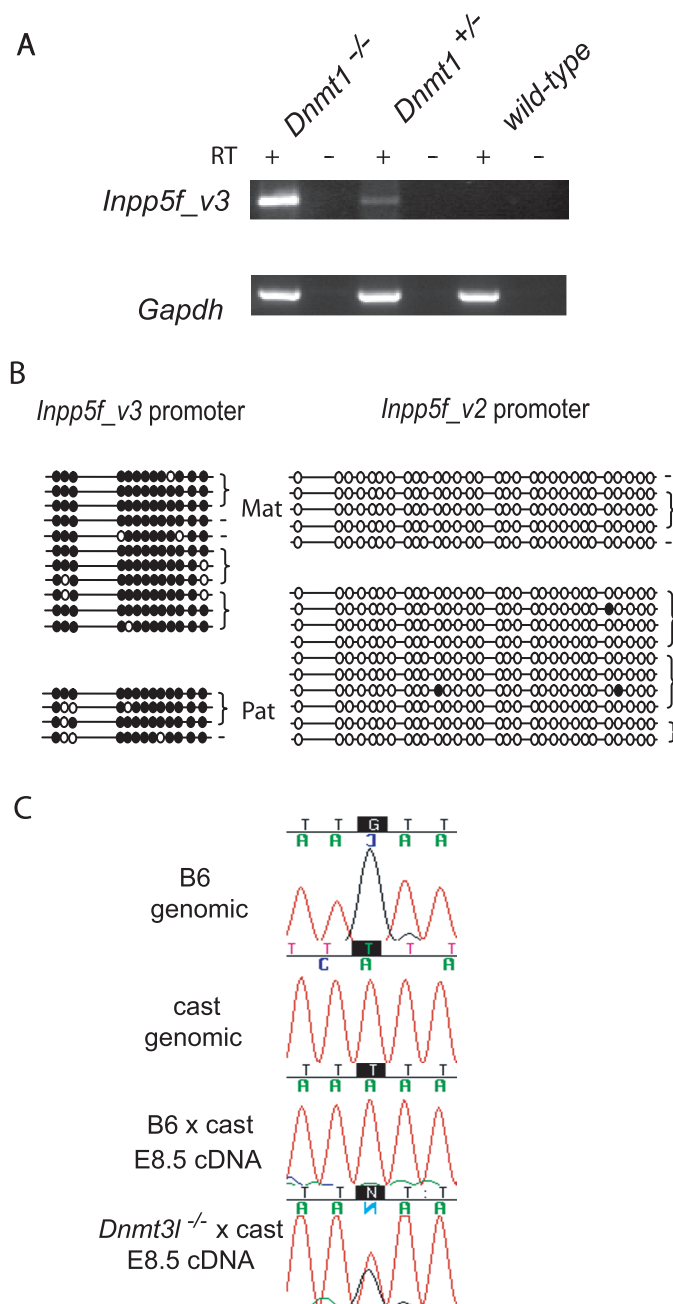
**Figure 5.** Tissue-specific methylation differences at the *Inpp5f\_v3* promoter in an expressing and non-expressing tissue. The overall percentage of methylated CpGs on maternally (Mat) and paternally (Pat) derived alleles is indicated in each case. Strands derived from the same PCR amplification are connected by brace symbols to the right of the figure. (A) Neonatal brain and (B) Neonatal heart.

parental origin is located at the *Inpp5f\_v2* promoter, and that the onset of imprinted *Inpp5f\_v3* expression is likely to be a secondary event occurring after E8.5.

## DISCUSSION

Imprinted genes commonly occur in clusters, and genes within these clusters are coordinately regulated by one or a few DNA elements termed imprinting control regions [ICRs; (25)]. One property shared by all eight ICRs that have been defined by gene targeting, (*Igf2*, *Kcnq1*, *Igf2r*, *Snrpn*, *Gnas1a*, *Nespas*, *Rasgrf1*, *Dlk1*) is the ability to undergo differential CpG methylation during gametogenesis, and subsequently maintain these allele-specific differences after fertilization during somatic development.

The CpG island at the promoter of *Inpp5f\_v2* overlaps a retrotransposed ORF with homology to the X-linked *Tmem114a* gene (10). A detailed methylation analysis of this region revealed that the allele-specific markings, which are established during gametogenesis, are maintained throughout embryonic development (Figure 4). Furthermore, embryos derived from *Dnmt3l*<sup>-/-</sup> mothers showed activation of the normally silent maternally derived allele (Figure 6C). These observations suggest that the CpG island with homology to the X chromosome is integral to the imprinting mechanism at this locus.



**Figure 6.** Disrupted expression of *Inpp5f\_v2* and *Inpp5f\_v3* in *Dnmt1* and *Dnmt3l* mutants. (A) Qualitative RT-PCR expression assay for *Inpp5f\_v3* in E8 embryos carrying null mutations in *Dnmt1*. *Inpp5f\_v3* was amplified for 34 cycles, *Gapdh* was amplified for 30 cycles. Parallel amplifications were performed using cDNA generated in the presence (+ RT) or absence (-RT) of reverse transcriptase. (B) Allele-specific CpG methylation at the two promoter regions in E8.5 embryos derived from *Dnmt3l*<sup>-/-</sup> mothers (on a *Mus musculus domesticus* background) and wild-type cast fathers. (C) Allele-specific RT-PCR sequencing assay for *Inpp5f\_v2* in E8.5 embryos derived from mothers carrying homozygous null mutations in the *Dnmt3l* gene, on a *Mus domesticus* background. The maternal strain or genotype is given first in the hybrid crosses.

Despite also undergoing differential methylation during gametogenesis, the *Inpp5f\_v3* promoter undergoes *de novo* methylation on the paternally derived allele between E3.5 and E8.5 (Figure 4B and C). The genome undergoes



extensive epigenetic reprogramming during this period (26), including the acquisition of methylation at Alu elements in primates (27,28). Murine B1 repeats are homologous to primate Alu elements; both families originated from 7SL, the RNA component of the signal recognition particle (29). Thus, the inverted B1 elements situated <100 bp upstream of the CpG-rich repeats (Figure 3A) may attract methylation to the surrounding region at the implantation stage. A similar B1 repeat upstream of the murine *Aprt* gene is capable of attracting *de novo* methylation to surrounding sequences in a cell culture system (30).

The activation of *Inpp5f\_v3* in *Dnmt1*<sup>-/-</sup> embryos shows that maintenance DNA methylation is required for *Inpp5f\_v3* silencing *in vivo* (Figure 6A). The proximity of the promoter region to tandem B1 elements suggests that the SINEs may contribute to the developmental expression profile of this gene. This provides an attractive explanation for the paucity of murine B1 and primate Alu elements found in the vicinity of imprinted promoters (12,13). Appropriate DNA methylation at imprinted loci is essential for embryonic viability (6), but may be disrupted by SINE insertions with potentially deleterious consequences. In this regard, *Inpp5f\_v3* can be viewed as exceptional. Here, the epigenetic regulation of the repeat-rich sequences may be utilized by the host organism to achieve the appropriate developmental expression pattern for *Inpp5f\_v3*. Alternatively, the imprinting and tissue-specific expression of *Inpp5f\_v3* may serve no intrinsic function but may occur as a downstream consequence of paternal-allele-specific *Inpp5f\_v2* expression, which is highest in neural tissues (21).

Despite biallelic hypermethylation at E8.5, the paternal *Inpp5f\_v3* promoter subsequently loses methylation and is expressed while the maternal allele remains hypermethylated and silent (Figures 2C and 4D). Demethylation is more pronounced in the brain than in non-expressing heart tissue, suggesting a model in which active chromatin 'spreads' from the *Inpp5f\_v2* promoter region which is active primarily in neural tissues (Figure 2B). This could occur due to the binding of transcription factors, which is known to cause demethylation of local sequences in dividing cells (31,32). To our knowledge, *Inpp5f\_v3* is the first example in mammals of an imprinted gene that undergoes allele-specific demethylation during somatic development.

The ORF of *Inpp5f\_v2* is one of a group of at least four imprinted retrogenes that originated from X-linked source genes (10). The data presented here show that the promoter region exhibits the epigenetic characteristics of an ICR, which could control the imprinting of both the *Inpp5f\_v2* and *Inpp5f\_v3* transcripts. *U2a1-rs1* and *Mcts2*, two other X-to-autosome retrogenes, also physically overlap gDMRs that are likely to control the imprinting of adjacent genes (10,33). We have recently identified additional imprinted transcripts at the fourth locus: *Nap115* (Wood *et al.*, unpublished data). Thus, all of the four known gDMRs that overlap X-to-autosome retrogenes are associated with multiple imprinted transcripts. These observations, combined with recent findings at the *Peg10* locus (9), suggest that the (retro)transposition of

elements from the X chromosome is one mechanism by which mammalian imprinted domains were generated during evolution.

## SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

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